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CHAPTER 4.3

# Microdialysis in the study of behavior reinforcement and inhibition

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**Abstract:** Brain microdialysis has been a valuable technique in the neuroscience field for more than 20 years. In vivo microdialysis in freely moving rats allows measurement of neurotransmitter release in response to ongoing behaviors. In this chapter we review findings using microdialysis in the study of behavior reinforcement and inhibition. This literature leads to the development of the dopamine hypothesis of reward and the cholinergic hypothesis of aversion and their underlying neural circuitry. Within the context of natural rewards, we discuss many of the key findings using microdialysis in the nucleus accumbens, ventral tegmental area and hypothalamus to study feeding, water intake, and mating. Artificial rewards, such as intracranial self-stimulation and drug reward, are also reviewed. Finally, data are summarized that suggest a natural reward, sugar, may take on behavioral and neurochemical properties of an artificial reward, such as a drug of abuse, under certain conditions

## I. Introduction

### I.A. Brain microdialysis

The microdialysis technique is a valuable tool in basic and clinical research. It has been extensively and successfully used, particularly in the neuroscience field, for almost three decades (Mark et al., 1991; Ungerstedt, 1991; Westerink, 1995; Muller, 2002; Bourne, 2003; Plock and Kloft, 2005). The popularity of this technique is due to its advantages with respect to other in vivo sampling techniques. It permits recovery of endogenous and exogenous substances from the brain or body, or infusion of drugs through the microdialysis probe (i.e., reverse microdialysis). Dialysis occurs by solute exchange through the porous membrane of the

probe following chemical gradients. The collected amount of the analytes in the dialysate is proportional to what is released near the probe (Segovia et al., 1986; Schwartz et al., 1990). Since the dialysates are pure protein-free ultrafiltrates, they do not suffer enzymatic degradation and can be chemically analyzed without pretreatments. Thus, it is possible to continuously sample from the extracellular environment in virtually all living tissues and organs of sufficient size. The analytes are typically analyzed by high-performance liquid chromatography (HPLC). Coupling microdialysis with improved analytical techniques such as capillary electrophoresis or mass spectrometry allows for the study of chemical changes in very short periods and in very small sample volumes (see Chapter 3.4).

The main application of microdialysis has traditionally been the study of chemicals in the brain. Before the invention of microdialysis,

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neurotransmission studies were based on techniques such as push–pull perfusion and tissue homogenates (Korf, 1986; Myers et al., 1998; Kottogoda et al., 2002; Rui and Lebaron, 2005). Microdialysis initially studied chemical changes in discrete brain areas of anesthetized rats (Hernandez et al., 1983; Segovia et al., 1986), but rapid improvements in the technique soon made it possible to sample from freely behaving animals (Zetterstrom and Ungerstedt, 1984; Hernandez et al., 1986; Carboni et al., 1989; Westerink, 1995; Fillenz, 2005). Microdialysis in freely behaving animals is ideal for studying the relationship between particular chemical messengers and changes in ongoing behaviors, such as feeding (Hoebel et al., 1989; Hernandez et al., 1991; Meguid et al., 1996; Bassareo and Di Chiara, 1999a, b; Rouch et al., 1999; Smith, 2004; Rada et al., 2005), drinking (Yoshida et al., 1992; Tanaka et al., 2004; Molander et al., 2005), mating (Pfaus et al., 1990; Dominguez and Hull, 2005), exercising (Meeusen et al., 2001), cognitive processes (Pepeu and Giovannini, 2004), pain (Stiller et al., 2003), intracranial self-stimulation (ICSS) (Hernandez and Hoebel, 1988a; You et al., 2001), drug self-administration (Hurd et al., 1999; Ranaldi et al., 1999), and in neuropsychiatric animal models (Hernandez et al., 1990, 1991; Hoebel et al., 1992; Joseph et al., 2003; Invernizzi and Garattini, 2004; Wilcox et al., 2005).

A Medline search indicates there have been more than 10,000 *microdialysis* publications in the last 25 years, with no indication of a decline in its use. So far, 71% of microdialysis studies are in *rats*, and of those 74% are in the *brain*. Behavioral studies represent 18% of all microdialysis studies, and reward studies correspond to about 10% of those, almost all of which have been conducted in rats.

### ***I.B. The concept of neural reward and aversion processes***

This chapter will review some of the findings that the microdialysis technique has offered in the field of behavior reinforcement and its inhibition. Pavlov (1927) used the term “reinforcement” to

refer to the strengthening of the association between an unconditioned stimulus and a conditioned stimulus that results when the two are paired. Skinner (1938) defined a “reinforcer” as a stimulus administered following a correct, arbitrarily chosen, response that increases the probability of occurrence of the response. The terms reinforcement and reward will be used interchangeably in this chapter and will include both positive and negative reinforcement (i.e., behavior to get, or get rid of, the reinforcer). Primary reinforcers, such as food, water, and sex, have an inherited role in proliferating the survival of the animal or the species, while secondary reinforcers, such as associated environmental stimuli, are learned.

Aversion, or behavior inhibition, will be considered as the opposite of reward in that it is a process that suppresses, instead of increasing, a behavior. This type of response can also be a lifesaver. For example, a conditioned taste aversion prevents an animal from eating a food that has previously been associated with sickness. Normally, once the consummatory phase of a natural behavior has been satisfied an inhibition of the behavior occurs. This behavior inhibition, with regard to feeding, is defined as the satiation process leading to satiety, and although the animal is averse to further eating, the state is generally described as pleasant.

In this chapter, we will review results that suggest the dopamine (DA) system in the nucleus accumbens (NAc) is part of a reinforcement system. DA release can activate an animal and contribute to a desired state. Cholinergic interneurons may counteract DA and play a role in behavior inhibition. Thus depending on the balance of neurotransmitter functions and the circuits in which they are embedded, an animal will either increase or decrease its response rate or response force. We hypothesize that high extracellular DA reinforces behavior, be it approach or escape behavior. If acetylcholine (ACh) also rises the approach behavior becomes inhibited and satiation ensues. Escape behavior lowers ACh. Low extracellular DA coupled with chronically increased ACh release depresses an animal and may lead to a condition

1 variously described as immobility, helplessness, or  
 2 despair.

### 5 *I.C. Anatomical substrates of reward*

7 The neural circuitry involved in reward and aver-  
 8 sion is part of brain systems engaged in emotions.  
 9 The limbic system was originally conceived as a  
 10 series of interconnected brain areas that played an  
 11 important role in the acquisition, storage, and ex-  
 12 pression of emotion, and was described as a circuit  
 13 between hypothalamus and the cerebral cortex  
 14 (Papez, 1995), and later expanded to include other  
 15 structures such as the amygdala (AMYG), hippo-  
 16 campus (HIPPO), and NAc. From early studies by  
 17 Mogenson and collaborators, it was concluded  
 18 that the NAc was a structure that linked “emo-  
 19 tions to action” (Mogenson et al., 1980). In the  
 20 1950s, Olds and Milner (1954) serendipitously dis-  
 21 covered that rats would self-administer electrical  
 22 pulses directly into many of the sites that coincided  
 23 with areas of the limbic system, suggesting that the  
 24 brain had specialized “centers” for reward. Many  
 25 researchers contributed to the effort of tracing the  
 26 “reward pathway” starting with the circuit from  
 27 the lateral hypothalamus (LH) to the ventral teg-  
 28 mental area (VTA) and the NAc as proposed by  
 29 Wise and Hoffman (1992). As can be seen in Fig.  
 30 1, the neural circuitry of reward comprised at least  
 31 two loops. The first loop includes the LH and  
 32 connects to structures in the hindbrain, returning  
 33 to the NAc and then back to the hypothalamus  
 34 directly and indirectly through the ventral pallid-  
 35 um (VP) (Leibowitz and Hoebel, 2004; Kelley et  
 36 al., 2005). Others have described links from mid-  
 37 brain cholinergic cells to the VTA (Yeomans et al.,  
 38 2001), and a glutamatergic/orexin path directly  
 39 from the LH to the VTA and to the NAc (Harris et  
 40 al., 2005). A second “loop” is composed of cortical  
 41 glutamatergic inputs to the NAc with connections  
 42 from the NAc to the VP, then to the mediodorsal  
 43 thalamus projecting back to the prefrontal cortex  
 44 (PFC). This second loop, which is actually a series  
 45 of concentric loops or a spiral, receives important  
 46 afferents from the AMYG and HIPPO (McGinty,  
 47 1999; Napier and Mitrovic, 1999; McFarland et  
 al., 2004). This cortical–subcortical loop involving

the basal ganglia motor system also branches off in  
 the thalamus to connect with the primary motor  
 system. It is evident from the perspective of Fig. 1,  
 following Mogenson’s “reward circuitry,” theory  
 that the NAc is at one of the intersections between  
 limbic and motor systems. Another classic view  
 has received renewed attention with modern evi-  
 dence that the LH connects reciprocally with the  
 cortex (Rolls, 1984; Oomura, 1988). Neuroanato-  
 mists later described the extended amygdala as in-  
 terconnecting structures that shared strong  
 histological homologies, including the AMYG,  
 stria terminalis, substantia innominata, and the  
 NAc (de Olmos and Heimer, 1999). This designa-  
 tion clearly differentiated the NAc shell, as part  
 of the extended AMYG, from the NAc core and  
 dorsal striatum (STR) (Zahm et al., 1996; Cadoni  
 and Di Chiara, 1999; Weiner, 2003). Efforts to  
 differentiate the specific functions of the NAc shell  
 vs. core have led to very interesting hypotheses of  
 Pavlovian-instrumental transfer, learning, motiva-  
 tion, and habit formation, which are reviewed  
 elsewhere (Robbins et al., 1989; Kalivas and  
 Nakamura, 1999; Di Chiara, 2002).

### 27 *I.D. Dopamine hypothesis of reward*

29 Initial studies in the early 1970s showed that spe-  
 30 cific lesions of the DA projections from the mid-  
 31 brain to the STR produced deficits in feeding and  
 32 drinking, resembling the “lateral hypothalamic  
 33 syndrome” characterized by aphagia and adipsia  
 34 (Teitelbaum and Epstein, 1962; Ungerstedt, 1970,  
 35 1971). The deficit in feeding and drinking occurred  
 36 without somatosensory impairment but rather as a  
 37 form of sensory neglect (Zigmond and Stricker,  
 38 1972; Marshall and Teitelbaum, 1974; Lindholm et  
 39 al., 1975). It was suggested that the dopaminergic  
 40 system in the medial forebrain bundle was prob-  
 41 ably damaged as the ascending fibers ran through  
 42 the far-lateral LH. Studies by Wise and colleagues  
 43 and others have demonstrated that neuroleptics  
 44 (DA receptor antagonists) attenuate the expres-  
 45 sion of various behaviors such as feeding, drink-  
 46 ing, ICSS, and drug self-administration (Wise,  
 47 1978; Xenakis and Scalfani, 1981; Bailey et al.,  
 1986; Ettenberg and Camp, 1986; Ettenberg, 1989;

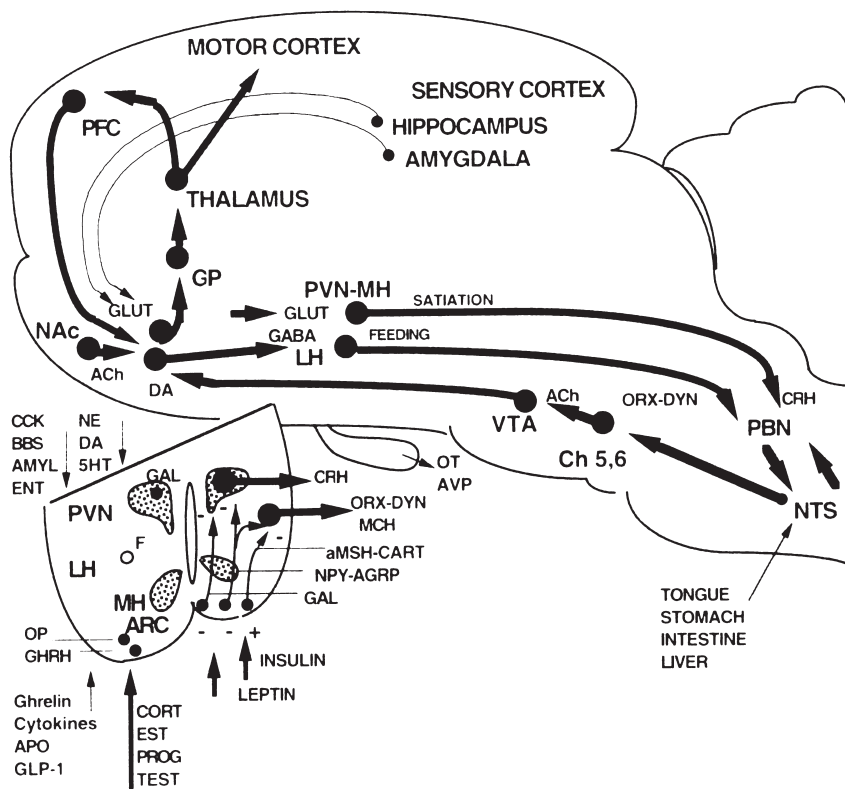


Fig. 1. Diagram of the reward circuitry showing the convergence in the NAc of the cortical and subcortical loops.

Wise and Rompre, 1989; Horvitz et al., 1993; Samson and Chappell, 2004; Xi et al., 2005a, b). Microdialysis studies have given further support to these behavioral findings by showing that natural reinforcers, as well as artificial reinforcers such as ICSS and drugs of abuse increase extracellular DA in the NAc (Hernandez and Hoebel, 1988b; Nakahara et al., 1989a, b; Wise et al., 1995a; Rinaldi et al., 1999). Electrophysiological studies in monkeys have emphasized the role of dopaminergic neurons in the animal's capacity to predict the occurrence of a novel event and reward learning (Schultz, 1997, 1998a, b; Tobler et al., 2005). Berridge and Robinson (1998) also suggest that DA may be involved in "incentive salience" and emphasize this role in motivation. Their conclusions are based on several experiments using "taste reactivity" to measure affective reactions suggesting that DA-depleted rats have normal hedonic reactions and associative learning (Berridge

and Robinson, 1998). Moreover, DA blockers do not seem to block oral approach reflexes, nor do specific DA agonists potentiate such measures, but they do alter the incentive value of the reward (Pecina et al., 1997, 2003; Berridge and Robinson, 1998). Recently, in DA-deficient mice, it was demonstrated that DA was not necessary for learning or liking, but it was necessary for seeking of the reward (Robinson et al., 2005), which could be reinstated by localized restoration of DA function with viral vectors (Szczycka et al., 2001). Most recently, brain imaging studies have been used to detect the active parts of the brain during various aspects of reinforcement, with attention drawn to changes in DA receptor function during the cognitive aspects of reinforcement (Kalivas and Volkow, 2005).

Stressful events and aversive stimuli can also increase DA levels in the NAc (Abercrombie et al., 1989; Keefe et al., 1993; Salamone, 1994;

1 Salamone et al., 1997; Rada et al., 1998b). On the  
 3 surface this seems to argue against any theory that  
 5 DA mediates reward, but on reflection, there is  
 7 more to reward than simple pleasure. DA could  
 9 also be involved in the relief from stress and pain,  
 both in the motivation to achieve relief and the  
 reward of achieving it. This fits with the theory  
 that DA has a role in salience and in negative re-  
 inforcement as well as positive reinforcement.

11 None of these discoveries rule out the possibility  
 13 that DA activates a pleasure response at some  
 15 point in the circuit, including mu-opioid receptor  
 “hot spots” in the NAc for “liking” a taste (Pecina  
 and Berridge, 2005). The opioid peptide enkepha-  
 lin in the NAc has been related to reward and can  
 activate both mu and delta receptors to increase  
 the release of DA (Di Chiara and Imperato, 1988;  
 Bals-Kubik et al., 1989). Opiates are involved in  
 eliciting feeding in the NAc (Kelley et al., 2000) as  
 well as in many other limbic system sites (Levine  
 and Billington, 2004). Moreover, it was shown in  
 1980, in one of the first local self-administration  
 studies, that rats will self-administer morphine di-  
 rectly into the NAc (Olds, 1982).

25 Microdialysis experiments have encountered all  
 27 of these interesting dimensions to the study of  
 29 DA’s role in reward. DA increases in the NAc shell  
 during an animal’s first exposure to a novel food,  
 and this response disappears at a second meal even  
 though the animal consumes as much as the first  
 time (Bassareo and Di Chiara, 1999b; Bassareo et  
 al., 2002; Rada et al., 2005). The DA response can  
 be reinstated by food deprivation (Bassareo and  
 Di Chiara, 1999b) or an animal can learn to re-  
 store it, and obtain a DA surge every day, by binge  
 eating (Rada et al., 2005).

### 39 *I.E. Cholinergic hypothesis of aversion*

41 Microdialysis has also contributed to a theory of  
 43 “aversion” or behavior inhibition. The details are  
 given in later sections of this review under the  
 topics of feeding satiation, mating satiation, ICSS  
 escape and drug withdrawal. In brief, we find that  
 45 ACh is released in the NAc in a variety of situ-  
 47 ations that all have behavior inhibition as a com-  
 mon feature. Extracellular ACh rises toward the

end of a meal (Mark et al., 1992; Rada et al., 2005)  
 and is also elevated after experiencing the forced  
 swim test and thus may contribute to behavioral  
 depression (Chau et al., 2001). Acetylcholine also  
 increases in the NAc during aversive hypothalamic  
 stimulation, and most telling, extracellular ACh  
 levels decrease when the animal performs stimula-  
 tion–escape responses (Rada and Hoebel, 2001).  
 Evidence will be presented for the general principle  
 that accumbens ACh is relatively high compared  
 with DA during withdrawal from addictive sub-  
 stances (Rada et al., 1996, 2004; Colantuoni et al.,  
 2002; Rada and Hoebel, 2005). Since withdrawal is  
 an aversive condition that results in many behav-  
 ioral signs of distress, including anxiety and de-  
 pression, one can surmise that relatively high levels  
 of ACh in the NAc can enhance a circuit that can  
 cause either behavior inhibition or aversion or  
 both.

## II. Natural rewards

Rewards can be classified as nondrug rewards or  
 drug rewards (Di Chiara, 2002), or as natural and  
 artificial rewards. We will classify natural rewards  
 as those reinforcers studied in natural behaviors  
 (feeding, thirst, and mating) and artificial rewards  
 as drug reward and ICSS. Artificial rewards act via  
 natural pathways but have magnified responses.  
 We will focus first on natural rewards and later  
 show how they too can become magnified, thereby  
 blurring the distinction between nondrug and drug  
 rewards.

### *II.A. Feeding*

Of all the natural rewards under investigation,  
 feeding behavior is the most studied using the mi-  
 crodialysis technique. The study of ingestive be-  
 havior has progressed substantially during the last  
 decade with the discovery of new peptides involved  
 in feeding regulation and the development of new  
 animal models of feeding disorders (see compre-  
 hensive reviews by Berthoud, 2004; Leibowitz and  
 Hoebel, 2004). In the present chapter, we will fo-  
 cus specifically on findings using brain



1 microdialysis that have impacted the ingestive behavior field.

3 Most research on feeding behavior using brain  
5 microdialysis has concentrated on the hypothalamus and NAc. Fig. 1 shows a way in which  
7 these areas are linked (also see Rolls, 1994; Berthoud, 2000). The hypothalamus has maintained  
9 its early start in feeding research as an area clearly involved in feeding initiation and satiation, and a  
11 place where most feeding neuropeptides act (Leibowitz and Wortley, 2004). Circuits in the hypothalamus  
13 can foster foraging for one macronutrient or another (Leibowitz and Wortley, 2004) and are under the control of hormones that  
15 signal nutrient stores in the body (Leibowitz and Hoebel, 2004; Strader and Woods, 2005). The  
17 NAc is a terminal field of the mesolimbic dopaminergic system involved in hedonic and motivational  
19 aspects of feeding as discussed above. Many other limbic areas are also involved, but  
21 these two sites capture the basic subcortical functions of sensory input, physiological modulation  
23 and motor output. Higher structures project to limbic sites with information from learning and  
25 memory stores (Rolls, 2000). One can start anywhere in these wonderful circuits. Given the topic  
27 of this chapter, we start with the NAc.

## 29 *II.A.1. NAc microdialysis during feeding*

31 *II.A.1.a. Dopamine in the NAc.* Dopamine is the  
33 most studied neurotransmitter in relation to natural rewards. As mentioned briefly in the Introduction,  
35 lesion of the LH is characterized by aphagia and adipsia (Anand and Brobeck, 1951a, b;  
37 Teitelbaum and Epstein, 1962), and this syndrome was due in part to damage incurred by dopaminergic  
39 projections to the STR (Ungerstedt, 1970, 1971). This spurred the study of a possible  
41 role for DA in an animal's reaction to natural rewards. DA antagonists injected locally into the  
43 accumbens increase operant responding for food, which then gradually declines suggesting a loss of  
45 reward and not simply motor impairment (Wise, 1978). There is also a loss of reaction to the taste of  
47 sugar (Schneider et al., 1986). Microdialysis studies have shown that feeding releases DA in the  
NAc (Hernandez and Hoebel, 1988a, b;

1 Radhakishun et al., 1988; Hoebel et al., 1989; Westerink et al., 1994). An initial use of microdialysis  
3 to understand if DA in the NAc is involved in conditioning was carried out by Mark et al.  
5 (1991), showing that oral infusion of a palatable food, saccharin, increased DA release in the NAc,  
7 and the opposite response was observed if the saccharin had been associated with sickness in a  
9 conditioned taste aversion paradigm.

11 Di Chiara and colleagues (Bassareo and Di Chiara, 1997, 1999b; Tanda and Di Chiara, 1998)  
13 find that a novel, unconditioned, palatable food stimuli (Fonzies) releases DA only during the first  
15 experience, and this effect habituates on the second exposure, suggesting that DA in the NAc is  
17 involved in acquisition rather than maintenance of incentive motivation (Bassareo and Di Chiara,  
19 1997). The first experience with a novel food raises DA selectively in the shell and not the core of the  
21 NAc (Tanda and Di Chiara, 1998; Bassareo and Di Chiara, 1999a), and this increase is dependent  
23 on stimulation of mu-opioid receptors located in the VTA (Tanda and Di Chiara, 1998). Similar  
25 results have been found using sucrose. Ingestion of a 10% sucrose solution releases DA in the NAc  
27 shell during the first exposure, but less on a second exposure 24 h or 21 days later, again suggesting  
29 that DA release in the NAc shell depends on the novelty of the stimulus (Rada et al., 2005). DA  
31 release is proportional to sucrose concentration (Hajnal et al., 2004), and under certain feeding  
33 conditions DA can be released with a palatable food time after time, as discussed in Section III of  
this chapter (Rada et al., 2005).

35 Postingestional factors can influence DA release in the NAc. In a conditioned taste preference  
37 paradigm, a neutral taste that was previously associated with infusion of a highly caloric solution into  
39 the stomach can increase DA levels in the NAc (Mark et al., 1994), suggesting that the DA increase  
41 observed when rats drink sucrose could be due, in part, to prior experience with its caloric  
43 content (Hajnal and Norgren, 2001; Rada et al., 2005). An alternative explanation could be that  
45 DA increases in the NAc shell as a consequence of the orosensory stimulation. In order to bypass  
47 most postingestional factors a gastric fistula can be implanted so that a liquid diet (e.g., sucrose) can

1 be drained out (Mook et al., 1988; Smith, 1998).  
 2 Sham-feeding confirms that the taste of sucrose  
 3 can release DA in the NAc (Hajnal et al., 2004;  
 4 Avena et al., in press(b)), thereby corroborating  
 5 the effect of saccharin (Mark et al., 1991). To-  
 6 gether, these results demonstrate that DA release  
 7 in the NAc can vary with orosensory stimulation,  
 8 postingestive factors, and the presence of a novel  
 9 palatable food.

11 *II.A.I.b. Acetylcholine in the nucleus accumbens*  
 12 (*behavior inhibition*). Acetylcholine in the STR  
 13 and accumbens is released from interneurons that  
 14 represent 5% of the cellular population (Bolam et  
 15 al., 1984; Meredith et al., 1989). Some clues as to  
 16 the effects of cholinergic interneurons on reward  
 17 mechanisms can be deduced from Parkinson's dis-  
 18 ease. These patients have destruction of DA neu-  
 19 rons in the basal ganglia and present symptoms  
 20 such as bradikinesia, rigidity, and abnormal move-  
 21 ments (dorsal STR), and also an anhedonic state  
 22 (ventral STR) (Fibiger, 1984; Isella et al., 2003;  
 23 Lemke et al., 2005) including weight loss (Chen et  
 24 al., 2003; Palhagen et al., 2005; Tuite et al., 2005).  
 25 A general hypothesis suggests that normal func-  
 26 tioning of the basal ganglia depends on a balance  
 27 between DA/ACh in the STR (Grewald et al.,  
 28 1974). When the DA neurons degenerate, an im-  
 29 balance occurs, with a relative increase in ACh  
 30 release (Spehlmann and Stahl, 1976; Rodriguez-  
 31 Puertas et al., 1994). This led us to the theory that  
 32 ACh in the NAc could counteract DA's rewarding  
 33 signal and thus produce behavioral inhibition.

35 Few studies have investigated the role of ACh in  
 36 the NAc during feeding. Acetylcholine increases in  
 37 the accumbens at the end of the meal and probably  
 38 signals satiety (Mark et al., 1992). When neostig-  
 39 mine, an acetylcholinesterase inhibitor and indi-  
 40 rect cholinergic agonist, was infused through a  
 41 dialysis probe bilaterally in the NAc, a decrease in  
 42 food intake was observed (Mark et al., 1992).  
 43 Moreover, bilateral infusion of a muscarinic (M1)  
 44 receptor agonist (arecoline) shortens the time in-  
 45 terval to reach satiety (Rada et al., unpublished  
 46 data). Lesion of the cholinergic interneurons with  
 47 a specific neurotoxin makes rats eat more, al-  
 though they lose body weight (Galosi et al., 1997;  
 Hajnal et al., 2000). Further evidence that ACh

1 could be involved in behavioral inhibition comes  
 2 from conditioned taste aversion experiments, in  
 3 which an aversive taste increases ACh levels in the  
 4 NAc (Mark et al., 1995) and local injection of a  
 5 cholinergic agonist into the accumbens induces a  
 6 conditioned taste aversion (Taylor et al., 1992).  
 7 Finally, the hypothesis that ACh signals satiety  
 8 was tested using the sham-feeding paradigm. If  
 9 ACh signals satiety, then sham-fed rats should  
 10 consume large amounts of food without any  
 11 change in accumbens ACh. Indeed, rats that are  
 12 sham-fed drink large amounts of a liquid diet  
 13 (10% sucrose) and show no significant change in  
 14 ACh levels in the NAc during the meal. Evidently  
 15 postingestive signals are needed to activate ACh  
 16 interneurons during feeding behavior as can be  
 17 seen in Fig. 2 (Avena et al., in press).

19 Several anomalous behavioral results question  
 20 the idea that ACh serves as a behavior-inhibition  
 21 signal in the NAc, but there are alternative expla-  
 22 nations for these results. Carbachol, a nonspecific  
 23 cholinergic agonist, is self-administered directly  
 24 into the accumbens (Ikemoto et al., 1998). In a  
 25 feeding paradigm, local injection of the nonspecific  
 26 muscarinic antagonist, scopolamine, reduces lever  
 27 pressing for food and sucrose consumption and  
 28 increases locomotor activity (Pratt and Kelley,  
 29 2004; Kelley et al., 2005; Pratt and Kelley, 2005).  
 30 In these experiments a nonspecific agonist and an-  
 31 tagonist, were used, making the interpretation  
 32 difficult. For instance, brain microdialysis has  
 33 shown that local infusion of scopolamine into the  
 34 accumbens increases ACh release, probably by  
 35 antagonizing M2 presynaptic autoreceptors (Chau  
 36 et al., 1999, 2001), and local infusion of carbachol  
 37 not only decreases ACh release but also increases  
 38 DA release, possibly explaining why rats would  
 39 self-administer this drug (Fig. 3).

41 *II.A.I.c. Glutamate and GABA in the nucleus*  
 42 *accumbens*. Evidence has been accumulating for  
 43 roles of accumbens glutamate (GLU) in reward,  
 44 motivation and novelty (Saulskaya and Mikhail-  
 45 ova, 2004; Kalivas and Volkow, 2005). Local in-  
 46 jection in the NAc of an AMPA/kainate  
 47 antagonist is sufficient to stimulate feeding in sa-  
 tiated rats (Maldonado-Irizarry et al., 1995). This  
 was later corroborated by microdialysis, showing

QA 2

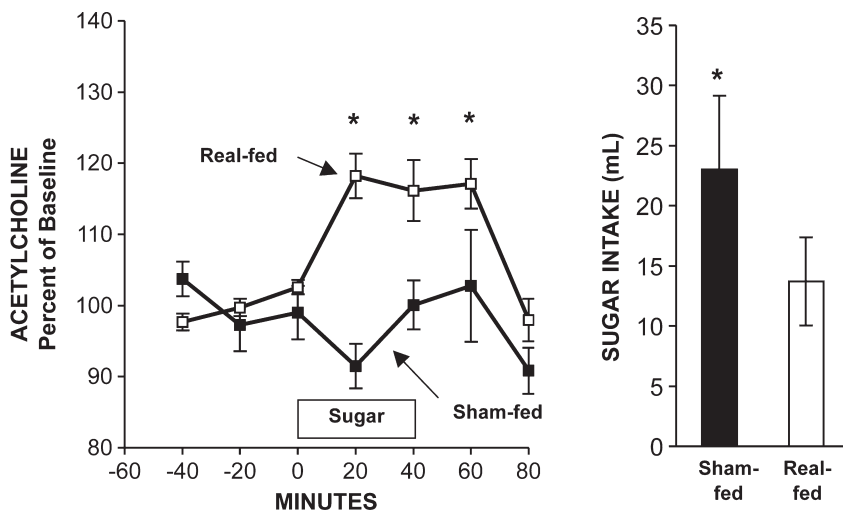


Fig. 2. Changes in extracellular levels of ACh in the NAc of sham-feeding and real-feeding rats. Acetylcholine only increased in real-feeding rats during sugar intake. Bar graphs indicate the amount of sugar consumed in the sham-feeding rats (black bars) compared with real-feeding rats (open bars). Asterisks indicate  $p < 0.05$ .

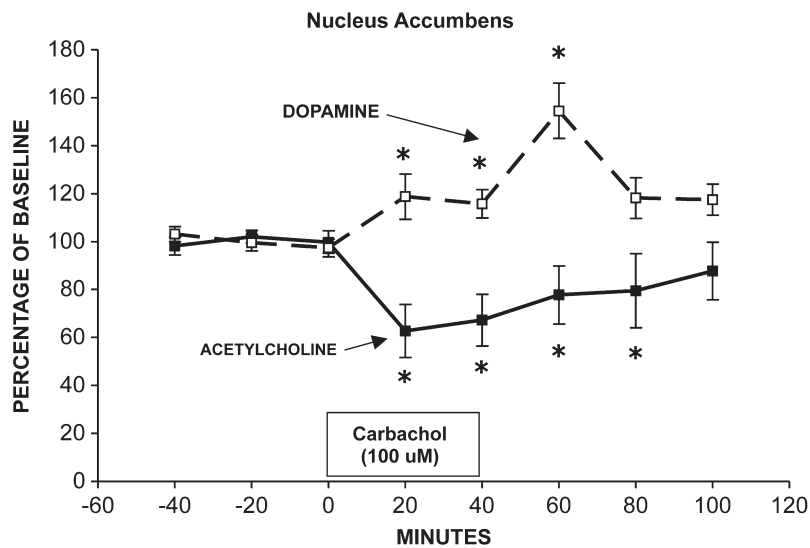


Fig. 3. A 40 min infusion of carbachol (100  $\mu$ M) in the accumbens by reverse microdialysis simultaneously decreased extracellular ACh and increased DA levels in the NAc. Asterisks indicate  $p < 0.05$ .

that free feeding in food-deprived rats significantly decreased GLU release in the NAc (Rada et al., 1997). Intraaccumbal injection of raclopride, a D2 antagonist, prevents the GLU decrease (Saulskaya and Mikhailova, 2002). Glutamate is released in the NAc following the presentation of an inedible

object when the rat expects food (Saulskaya and Mikhailova, 2002) and also during the presentation of a conditioned aversive stimuli (Saulskaya and Mikhailova, 2004), suggesting that GLU is released in the NAc when the motivational value of the reward is changed or aversive.



1 Several pharmacological studies show that local  
 2 injections of GABA agonists into the NAc increase  
 3 food intake (Reynolds and Berridge, 2002; Hanlon  
 4 et al., 2004); however, presently there are no  
 5 known microdialysis studies that have measured  
 6 the effect of feeding on GABA release.

#### 9 *II.A.2. Ventral tegmental area microdialysis during 10 feeding*

11 This brain area is of utmost importance since DA  
 12 neurons, forming the mesolimbic system, originate  
 13 in the VTA (Fallon and Moore, 1978). These DA  
 14 neurons are known to be under the influence of  
 15 other neurotransmitter systems including GLU  
 16 (Taber and Fibiger, 1997; Westerink et al., 1997;  
 17 Floresco et al., 2001; Harris and Aston-Jones,  
 18 2003), GABA (Cruz et al., 2004; Ye et al., 2004),  
 19 and the opioids (Cowen and Lawrence, 1999).

23 *II.A.2.a. Dopamine in the ventral tegmental  
 24 area.* Dopamine is not only released in its termi-  
 25 nal region, the NAc, but also in its dendritic area.  
 26 While the majority of research has focused on the  
 27 NAc, studies have shown that extracellular DA  
 28 increases in the VTA in response to opioids (Klite-  
 29 nick et al., 1992; Yoshida et al., 1993). With regard  
 30 to feeding, extracellular DA increases in the VTA  
 31 while an animal eats (Yoshida et al., 1992).

35 *II.A.2.b. Acetylcholine in the ventral tegmental  
 36 area.* The VTA receives cholinergic inputs from  
 37 the pedunculopontine nuclei (Woolf, 1991). Infu-  
 38 sion of nicotine directly into the VTA by reverse  
 39 microdialysis stimulates DA release in the NAc  
 40 (Nisell et al., 1994). Muscarinic receptors in the  
 41 VTA have also been shown to modulate feeding  
 42 and drinking behavior. For instance, local injec-  
 43 tion of a muscarinic antagonist suppresses feeding  
 44 and ICSS (Rada et al., 2000; Sharf and Ranaldi,  
 45 2006). Conversely, ICSS stimulates ACh release in  
 46 the VTA (Rada et al., 2000). In this brain area,  
 47 ACh seems to facilitate behavior by directly stim-  
 48 ulating DA neurons that release DA in the NAc  
 49 (Forster and Blaha, 2000; Yeomans et al., 2001).

1 *II.A.3. Hypothalamic microdialysis during feeding*  
 2 In the following paragraphs we will discuss some  
 3 of the neurochemical findings using brain micro-  
 4 dialysis to measure amines and amino acids in the  
 5 hypothalamic region that have important links to  
 6 the reward circuitry.

7 *II.A.3.a. Dopamine in the hypothalamus.* Dop-  
 8 amine in the hypothalamus is thought to play a very  
 9 different role in feeding than in the NAc. Early  
 10 studies suggested that DA in the LH might be in-  
 11 volved in the anorectic effect of amphetamine (Le-  
 12 ibowitz, 1975). This was later confirmed when a  
 13 local LH injection of sulpiride, a relatively specific  
 14 D2 antagonist, was sufficient to induce feeding and  
 15 drinking (Parada et al., 1988) and, with repeated  
 16 injections, obesity (Baptista, 1999). DA in the LH  
 17 may be essential in locomotion related to food and  
 18 water seeking (Parada et al., 1990). Microdialysis  
 19 studies have found that eating induces a significant  
 20 increase in DA levels in the LH, with no change  
 21 observed in rats fed intragastrically, suggesting  
 22 that oropharyngeal stimulation is important  
 23 (Yang et al., 1996). However, in this report sam-  
 24 ples were taken every 20 min, so it is difficult to  
 25 know whether the DA increase was signaling sati-  
 26 ety. DA increase is directly correlated with the  
 27 meal size and it has been suggested that obese  
 28 Zucker rats may have an inherently higher DA  
 29 “threshold” level for satiety in the LH (Yang and  
 30 Meguid, 1995). It is difficult to know if DA signals  
 31 satiety and obese rats have a higher threshold or  
 32 whether obese rats eat more because they release  
 33 more DA in the LH. Behavioral experiments sup-  
 34 port the DA satiety explanation since DA agonists  
 35 act as anorectics when injected into the LH (Le-  
 36 ibowitz, 1975). Moreover, rats self-administer a  
 37 DA antagonist directly into the LH and this re-  
 38 leases DA in the NAc, thus linking LH DA to the  
 39 inhibition of NAc DA reinforcement (Parada et  
 40 al., 1995).

41 The ventromedial hypothalamus (VMH) usually  
 42 has an opposing response to that observed in the  
 43 LH. The VMH sends GABAergic projections to  
 44 the LH that may inhibit feeding (Beverly and  
 45 Martin, 1989). The VMH also receives do-  
 46 paminergic inputs, which respond in the opposite  
 47 manner to the LH, with a decrease in extracellular

1 DA concentrations during a meal (Yang et al.,  
 3 1997), which depends directly on the size of the  
 5 meal (Meguid et al., 1997). The DA decrease in the  
 7 VMH depends in part on oropharyngeal stimula-  
 9 tion (Yang et al., 1997). In summary, DA release  
 11 in the hypothalamus is involved in feeding be-  
 13 havior and the LH and VMH probably have op-  
 15 posing dopaminergic functions. The source of DA  
 17 in the hypothalamus could be local DA cell clus-  
 19 ters as well as the mesolimbic system (Fuxe and  
 21 Ungerstedt, 1968).

23 *II.A.3.b. Norepinephrine in the hypo-*  
 25 *thalamus.* Extracellular norepinephrine (NE) in  
 27 the hypothalamus follows a circadian rhythm,  
 29 suggesting that it plays an important role in the  
 31 animal's overall state of arousal (Margules et al.,  
 33 1972; Jacobs and Chan, 1987; Stanley et al., 1989).  
 35 The earliest studies, and more precisely in the  
 37 paraventricular nucleus (PVN), can enhance food  
 39 intake (Grossman, 1960; Leibowitz, 1970, 1972).  
 41 Microdialysis studies found that NE is released in  
 43 the PVN at the beginning of the active feeding  
 45 period coinciding with the onset of the active cycle  
 47 of the rat (dark onset) (Hoebel et al., 1989; Stanley  
 et al., 1989; Mitome, 1994; Morien et al., 1995;  
 Tachibana et al., 2000, 2001). It was later found  
 that PVN NE increases in satiated rats during a  
 large meal at dark onset, and it also increases at  
 the start of the dark cycle in food-deprived rats  
 given a carbohydrate meal (Paez et al., 1993). In  
 rats maintained on a restricted schedule, NE levels  
 rise just before the meal (Mitome et al., 1994).

Further studies have looked at the effect of var-  
 ious drugs or peptides, which can modify ingestive  
 behavior, on hypothalamic NE. For instance, ga-  
 lanin (GAL) and neuropeptide-Y (NPY) are both  
 peptides that, if injected in the PVN, can induce  
 feeding (Kyrkouli et al., 1990), and also increase  
 NE levels in the PVN in rats if food is present  
 (Kyrkouli et al., 1992). If food is not present, GAL  
 still increases NE levels, but NPY decreases it  
 (Kyrkouli et al., 1992). Intraventricular injection  
 of NPY increases both food intake and NE in the  
 PVN (Matos et al., 1996). Alpha-adrenergic an-  
 tagonists are capable of blocking the GAL- but  
 not the NPY feeding response (Kyrkouli et al.,  
 1990). These results are consistent with previous

behavioral studies showing that GAL-induced eat-  
 ing is probably mediated through the NE system.

Anorectic drugs modify NE release in the hypo-  
 thalamus. For instance, systemic injection of phe-  
 nylpropranolamine, an alpha-1 adrenoceptor  
 agonist, suppresses food intake in rats and simul-  
 taneously decreases extracellular levels of NE in  
 the PVN (Davies et al., 1993). In contrast, the al-  
 pha-2 blocker, idazoxan, also suppresses food in-  
 take in the rat, but instead of the expected decrease  
 in NE levels in the PVN an increase occurs, which  
 may be mediated through a presynaptic auto-  
 receptor (Paez and Leibowitz, 1993). These results  
 illustrate the importance of microdialysis in rec-  
 ognizing possible pre- and postsynaptic mecha-  
 nisms of action for various peptides and drugs that  
 modulate ingestive behavior.

Few studies have looked at the effect of obesity  
 on NE levels in the hypothalamus, although it is  
 known that chronic infusions of NE into the VMH  
 or the PVN induce hyperphagia and obesity (Le-  
 ibowitz et al., 1984; Cincotta et al., 2000). One  
 model of obesity uses male offsprings of female  
 rats that were undernourished during the first two  
 trimesters of pregnancy or had been injected with  
 insulin during the third trimester (Jones and Fried-  
 man, 1982; Jones and Dayries, 1990). Microdial-  
 ysis of the medial hypothalamus in the obese  
 offspring showed a significant elevation in extra-  
 cellular NE levels compared with control rats  
 (Jones et al., 1995). Thus, NE may play a role in  
 feeding or body weight regulation in this model of  
 gestation-linked obesity.

*II.A.3.c. Histamine in the hypothalamus.* Phar-  
 macological studies have demonstrated that local  
 hypothalamic histamine modulates food intake  
 (Ookuma et al., 1989). This was confirmed by lo-  
 cally manipulating histamine levels in the PVN  
 and VMH using an inhibitor of the synthetic en-  
 zyme histidine decarboxylase. Experimentally de-  
 creasing histamine levels induces feeding, but only  
 at the start of the light cycle and only in the PVN  
 or VMH, not in the LH or dorsomedial hypo-  
 thalamus (Ookuma et al., 1993). Histamine may  
 modulate ingestive behavior in the hypothalamus  
 by interacting with the noradrenergic system. Lo-  
 cal injection of a histamine H1 receptor antagonist

1 increases feeding and extracellular NE, and this  
 2 effect is blocked by a specific alpha-2 adrenoceptor  
 3 antagonist (Kurose and Terashima, 1999).

5 *II.A.3.d. Serotonin in the hypothalamus.* Medial  
 6 hypothalamic (MH) injection of serotonin (5-HT)  
 7 or its agonists inhibit feeding behavior (Leibowitz,  
 8 1986; Leibowitz et al., 1987, 1988). Initial micro-  
 9 dialysis studies of 5-HT during a meal demon-  
 10 strated that 5-HT increased in the LH and also in  
 11 the medial hypothalamus in anticipation of a meal  
 12 when smelling food, as well as during the meal  
 13 (Schwartz et al., 1990), suggesting that 5-HT may  
 14 play a role in the response to food-related  
 15 appetitive stimuli and then contribute to satiety  
 16 when postingestional factors release CCK or re-  
 17 lated satiety signals. Later studies demonstrated  
 18 that 5-HT increases if the rat is given a carbohy-  
 19 drate meal, and this increase is detected 15 min  
 20 after the start of the meal, possibly contributing to  
 21 satiety. In this same report, it was found that a  
 22 protein or fat meal could decrease 5-HT levels  
 23 (Rouch et al., 1999). This may relate in part to  
 24 tryptophan uptake for 5-HT synthesis after a car-  
 25 bohydrate meal (Fernstrom and Wurtman, 1974).  
 26 Other researchers have found changes in hypo-  
 27 thalamic 5-HT levels in normal and obese rats  
 28 consistent with the theory that 5-HT can act as a  
 29 satiety signal (Mori et al., 1999; Fetissov et al.,  
 30 2000; De Fanti et al., 2001). Several groups have  
 31 investigated the effect of satiety peptides on hypo-  
 32 thalamic 5-HT release. Enterostatin or leptin can  
 33 increase extracellular 5-HT in the LH (Koizumi  
 34 and Kimura, 2002; Telles et al., 2003). Conversely,  
 35 Orosco and collaborators suggest that 5-HT  
 36 causes the release of insulin in the PVN-VMH re-  
 37 gion (Orosco et al., 2000). These microdialysis  
 38 studies point to a role for 5-HT in modulating  
 39 circuits that control food intake, including a  
 40 strong synergistic effect with postingestional sati-  
 41 ety factors.

43 *II.A.3.e. Acetylcholine in the hypothalamus.* In  
 44 the LH several studies have demonstrated the po-  
 45 tentiation of eating and drinking water following  
 46 local injection of a muscarinic agonist (Grossman,  
 47 1960; De Parada et al., 2000). However, so far  
 there seems to be no published microdialysis

studies of ACh release in the LH during feeding  
 behavior.

5 *II.A.3.f. Glutamate and GABA in the hypo-*  
 6 *thalamus.* Retrograde labeling shows that GLU  
 7 inputs to the LH originate in the frontal cortex,  
 8 AMYG, NAc, preoptic area, SN, VTA, par-  
 9 abraquial nuclei, and the nucleus of the solitary  
 10 tract (Duva et al., 2005). Glutamate has been  
 11 shown to induce feeding when injected locally in  
 12 the LH (Stanley et al., 1993a, b; Khan et al., 1999;  
 13 Duva et al., 2002). Moreover, injection of NMDA  
 14 into the LH induces eating without affecting lo-  
 15 comotion (Duva et al., 2001, 2002). Extracellular  
 16 levels of GLU in the LH increase at the beginning  
 17 of a meal and then decrease by the end of the meal  
 18 (Rada et al., 2003). It would seem that GLU in-  
 19 itiates eating as a fast-acting neurotransmitter, and  
 20 the maintenance of the behavior probably depends  
 21 on other neurotransmitters.

22 It has been suggested that there exists reciprocal  
 23 connections between the medial and LH. In the  
 24 medial hypothalamus GABA can induce eating  
 25 (Beverly and Martin, 1989, 1990). Furthermore,  
 26 following acute glucoprivation that would increase  
 27 appetite, extracellular GABA increases in the  
 28 VMH while an opposite response occurs in the  
 29 LH (Beverly et al., 1995). Early studies injecting a  
 30 GABA agonist into the LH showed an increase in  
 31 feeding, while antagonists did the opposite (Kelly  
 32 et al., 1977; Kelly and Grossman, 1979; Tsujii and  
 33 Bray, 1991). However, later studies suggest a de-  
 34 crease in feeding when a GABA agonist is locally  
 35 injected into the LH (Maldonado-Irizarry et al.,  
 36 1995), but an antagonist does not initiate feeding  
 37 (Stratford and Kelley, 1999). Monitoring GABA  
 38 in the LH every 30 s during a meal it was found  
 39 that it increased at the end of the meal, possibly  
 40 signaling satiety (Rada et al., 2003). This could  
 41 also explain the absence of response using antag-  
 42 onists in the LH to induce feeding, since GABA  
 43 levels might be too low before the meal for an  
 44 antagonist to show any effect at that time.

#### 45 *II.A.4. Hypothalamic accumbens connections*

46 There are both direct and indirect connections be-  
 47 tween the hypothalamus and the NAc (Kelley et

al., 2005). Microdialysis has provided evidence that these sites may interact. Most, but not all, neurotransmitters or peptides that promote feeding when injected into the hypothalamus also stimulate DA and decrease ACh release in the NAc, while satiating peptides do the opposite (Leibowitz and Hoebel, 2004). Injection of the cholinergic agonist, carbachol or the DA antagonist, sulpiride, into the LH both increases food intake and simultaneously increases DA release in the NAc (De Parada et al., 2000). Similarly, injection of NE or GAL into the PVN of the hypothalamus increases food consumption, which is correlated with a significant increase in DA and decrease in ACh in the NAc (Hajnal et al., 1997; Rada et al., 1998). Conversely, cholecystokinin (CCK) injected into the PVN decreases food ingestion while decreasing DA in the NAc (Helm et al., 2003). Cholecystokinin does not increase ACh unless it is injected simultaneously with 5-HT, which then produces a profound decrease in food intake along with increased ACh release (Helm et al., 2003). This suggests that under normal circumstances, postingestional factors would provide CCK release for interaction with 5-HT contributing to the decrease in DA and increase in ACh release in the NAc. In summary, as a general rule, the hypothalamus controls feeding behavior, in part, by modulating DA and ACh in the NAc (Hoebel et al., 1999).

An exception to the hypothalamic accumbens rule is NPY. Injection of this peptide into the PVN induces eating in the rat, however, no changes in extracellular DA or ACh were detected (Rada et al., 1998a), suggesting that NPY can induce feeding by some other mechanism, or requires other cofactors that were not present.

## II.B. Water intake

Compared with the abundant data on food intake, there are relatively few brain microdialysis studies on water intake. It has been reported that drinking releases DA in the NAc and VTA (Yoshida et al., 1992; Young et al., 1992). Injection of a D2 receptor antagonist in perifornical LH can induce drinking and increase DA in the NAc (Parada et

al., 1988, 1990). This activation of the DA mesolimbic system could mediate some of the rewarding aspects of drinking behavior.

Similar to the effect seen with food intake, ACh increases in the VTA during drinking, and blockade with muscarinic receptors in the same area inhibits water intake (Rada et al., 2000). In the LH, extracellular ACh also increases during drinking, and the exogenous administration of cholinergic drugs or D2 receptor blockers increases water intake (Puig de Parada et al., 1997). These microdialysis studies are in agreement with behavioral studies that show an increase in water intake following pharmacological manipulation of the cholinergic system with carbachol (Grossman, 1960).

Angiotensin II injections into the subfornical organ induce drinking (McKinley et al., 2001) and release NE in the PVN and LH, even when water is not available (Gerstberger et al., 1992). However, this NE increase is attenuated if rats are allowed to drink (Ushigome et al., 2002; Tanaka et al., 2003). These studies indicate that a hypothalamic component of the noradrenergic system participates in drinking behavior. Angiotensin injection in the lateral ventricle releases DA in the NAc (Jones, 1986). This effect is enhanced if the rat is allowed to drink in response to the injection (Hoebel et al., 1994).

## II.C. Mating

The NAc participates in the control of sexual behavior in the same way that it does with other natural reinforcers. Microdialysis in the NAc of male and female rats shows an elevation of extracellular DA levels during sexual behavior (Damsma et al., 1992; Mas et al., 1995; Becker et al., 2001). In the NAc of sexually active male rats DA levels increase when a receptive female rat is presented, and increase even more during copulation (Pfaus et al., 1990; Pleim et al., 1990). Damsma and colleagues have examined the effects of locomotion, exposure to a novel chamber, sex odors, and sexual activity on DA transmission in the NAc and STR (Damsma et al., 1992). The DA increase seen during copulation is greater following active



1 locomotion (wheel running) or exposure to novel  
 3 stimuli (mating chamber, fresh bedding, or soiled  
 5 bedding). This increase was more intense in the  
 7 NAc than the STR. Thus, neither novelty nor lo-  
 9 comotion can account for the increase of DA in  
 either area, suggesting that the anticipatory and  
 consummatory aspects of sexual behavior are nat-  
 urally occurring events in which reinforcement is  
 likely mediated by DA release in the NAc.

11 Dopamine in the NAc and STR is also increased  
 13 during sexual behavior in females, but only when  
 15 the female controls the time of intromissions  
 (Becker et al., 2001). The timing of copulatory  
 17 stimuli is critical for the magnitude of the increase  
 19 in accumbens DA. When intromissions are spaced  
 1–2 min apart and insemination would most likely  
 result in pregnancy, DA significantly increases in  
 the NAc. This increase is not a passive response to  
 coital stimuli or copulation-related motor activity,  
 and possibly reflects other qualitative information  
 about the copulatory stimuli.

23 The medial preoptic area (MPOA) is located at  
 25 the rostral end of the hypothalamus and is a crit-  
 27 ical integrative site for male sexual behavior in  
 29 most vertebrate species. DA agonists in MPOA  
 facilitate sexual behavior, while antagonists impair  
 copulation, genital reflexes and sexual motivation  
 (Mas et al., 1987; Bitran et al., 1988; Warner et al.,  
 1991; Dominguez and Hull, 2005). Microdialysis  
 in the MPOA has shown a DA increase during  
 appetitive (noncontact exposure to sexual stimuli  
 such as exposure to a receptive female) and con-  
 35 summatory (copulation) phases (Fumero et al.,  
 1994; Dominguez et al., 2001; Triemstra et al.,  
 2005). Recently, it was shown that GLU, by stim-  
 37 ulating nitric oxide, is responsible for this increase  
 in MPOA DA during copulation (Dominguez et  
 al., 2004).

39 Bilateral olfactory bulbectomy completely pre-  
 41 vents mating in male rodents; however, unilateral  
 43 bulbectomy does not. Microdialysis performed in  
 the MPOA of animals with unilateral bulbectomy  
 during mating shows that DA increases only in the  
 contralateral and not in the ipsilateral side, sug-  
 45 gesting that somatosensory cues alone are not  
 sufficient to release DA in the MPOA during sex-  
 47 ual behavior in the absence of chemosensory input  
 (Triemstra et al., 2005). Sexually excitatory

1 olfactory stimuli activate the medial AMYG  
 (MeA), which in turn projects to the bed nucleus  
 of the stria terminalis and the MPOA leading to  
 3 DA release and facilitation of copulation (Ko-  
 5 starczyk, 1986; Gomez and Newman, 1992; Dom-  
 7 inguez et al., 2001).

9 Microdialysis in the anterior LH area suggests  
 11 that 5-HT inhibits behaviors during the postcop-  
 13 ulatory phase of male sexual behavior (Lorrain et  
 15 al., 1999). Serotonin injection in the LH also in-  
 17 hibits basal and female-induced DA release in the  
 NAc. This suggests that the neural circuit pro-  
 moting sexual quiescence during the postejacula-  
 tory interval include serotonergic input to the LH,  
 which in turn inhibits DA release in the NAc. This  
 fact may have relevance for understanding the  
 sexual side effects common to antidepressants  
 medications (Rudkin et al., 2004).

### III. Artificial rewards

#### III.A. Intracranial self-stimulation

25 Olds and Milner (1954) discovered ICSS in the mid  
 27 1950s to produce positive reinforcement. Re-  
 29 search, using intracerebral microdialysis, has fo-  
 31 cused mainly on how ICSS modulates DA release  
 in the NAc. Most studies show that LH or VTA  
 ICSS releases DA in the NAc (Hernandez and  
 33 Hoebel, 1988a; Nakahara et al., 1989a, b; Phillips  
 et al., 1992; Fiorino et al., 1993; You et al., 1998,  
 2001). Similarly, electrical brain stimulation of the  
 PFC significantly increases DA release in the NAc  
 (Taber and Fibiger, 1995; You et al., 1998, 2001).  
 This rise in NAc DA is intensity dependent and  
 37 can be blocked with the excitatory amino acid an-  
 39 tagonist kynurenic acid injected either into the  
 VTA or directly into NAc, suggesting that the  
 neural signal engages the VTA (You et al., 1998).  
 41 In addition, like food reward, self-stimulation of  
 the LH also increases ACh release in the VTA and  
 43 an infusion of atropine, through reverse dialysis,  
 completely blocks self-stimulation confirming that  
 45 this cholinergic system in the hindbrain is involved  
 in activating the DA neurons in the VTA (Rada et  
 47 al., 2000; Sharf and Ranaldi, 2006).



1 As discussed in previous paragraphs, an electro- 1  
 3 lytic lesion of the LH produces aphagia and 3  
 5 adipsia. Conversely, electrical stimulation of the 5  
 7 LH induces feeding in satiated animals (Anand 7  
 9 and Brobeck, 1951a, b; Hoebel and Teitelbaum, 9  
 11 1962; Valenstein et al., 1968; Hoebel, 1976). Using 11  
 13 microdialysis to monitor DA levels in the NAc, an 13  
 increase in DA levels was demonstrated following 15  
 17 electrical stimulation of the perifornical LH or 17  
 19 eating (Hernandez and Hoebel, 1988a; Rada et al., 19  
 21 1998b). This suggests that ICSS and feeding share 21  
 23 common circuitry and that DA in the NAc could 23  
 be part of the signal.

25 Using hypothalamic sites where electrical stim- 25  
 27 ulation was either positively or negatively rein- 27  
 29 forcing, or both, DA was released in the NAc by 29  
 31 automatic stimulation, self-stimulation or stim- 31  
 33 ulation–escape responding. DA increases even dur- 33  
 35 ing stimulation escape using a MH site that was 35  
 purely aversive (Rada et al., 1998b). These results 37  
 37 confirm that DA in the NAc is not only involved in 37  
 39 positive reinforcement, but in negative reinforce- 39  
 41 ment as well. 41

43 As cited in the prior section on ACh in the NAc, 43  
 45 the aversive LH stimulation causes release of 45  
 47 accumbens ACh. Stimulation–escape responding 47  
 significantly decreases extracellular ACh levels 1  
 (Rada and Hoebel, 2001). This supports the the- 3  
 5 ory that elevated ACh in the accumbens is avers- 5  
 7 1998b). This suggests that ICSS and feeding share 7  
 9 1998b). This suggests that ICSS and feeding share 9  
 11 1998b). This suggests that ICSS and feeding share 11  
 13 1998b). This suggests that ICSS and feeding share 13  
 15 1998b). This suggests that ICSS and feeding share 15  
 17 1998b). This suggests that ICSS and feeding share 17  
 19 1998b). This suggests that ICSS and feeding share 19  
 21 1998b). This suggests that ICSS and feeding share 21  
 23 1998b). This suggests that ICSS and feeding share 23  
 25 1998b). This suggests that ICSS and feeding share 25  
 27 1998b). This suggests that ICSS and feeding share 27  
 29 1998b). This suggests that ICSS and feeding share 29  
 31 1998b). This suggests that ICSS and feeding share 31  
 33 1998b). This suggests that ICSS and feeding share 33  
 35 1998b). This suggests that ICSS and feeding share 35  
 37 1998b). This suggests that ICSS and feeding share 37  
 39 1998b). This suggests that ICSS and feeding share 39  
 41 1998b). This suggests that ICSS and feeding share 41  
 43 1998b). This suggests that ICSS and feeding share 43  
 45 1998b). This suggests that ICSS and feeding share 45  
 47 1998b). This suggests that ICSS and feeding share 47

### III.B. Drug reward

1 Almost all drugs abused by humans increase DA 1  
 3 in the NAc (Di Chiara and Imperato, 1988; Her- 3  
 5 nandez and Hoebel, 1988b; Pothos et al., 1991; 5  
 7 Rada et al., 1991a, 2001; Tanda et al., 1997; Di 7  
 9 Chiara, 1998; Koob et al., 1998; Hoebel et al., 9  
 11 1999) with the exception of benzodiazepines and 11  
 13 barbiturates (Masuzawa et al., 2003; Rada and 13  
 15 Hoebel, 2005). Withdrawal in contrast decreases 15  
 17 DA release in the NAc (Parsons et al., 1991; Pot- 17  
 19 hos et al., 1991; Weiss et al., 1992; Diana et al., 19  
 21 1993, Hildebrand et al., 1998; Rada et al., 2004) 21  
 23 and, in several cases, increases ACh release (Rada 23  
 25 et al., 1991b, 1996, 2001, 2004). Although the 25  
 27 2001, 2004). Although the 27  
 29 2001, 2004). Although the 29  
 31 2001, 2004). Although the 31  
 33 2001, 2004). Although the 33  
 35 2001, 2004). Although the 35  
 37 2001, 2004). Although the 37  
 39 2001, 2004). Although the 39  
 41 2001, 2004). Although the 41  
 43 2001, 2004). Although the 43  
 45 2001, 2004). Although the 45  
 47 2001, 2004). Although the 47

1 benzodiazepine, diazepam (Valium), does not re- 1  
 3 lease DA, its withdrawal does release ACh. On the 3  
 5 theory that relatively high extracellular ACh is 5  
 7 aversive, this could contribute to the use of di- 7  
 9 azepam for self-medication (Rada and Hoebel, 9  
 11 2005). 11  
 13 2005). 13  
 15 2005). 15  
 17 2005). 17  
 19 2005). 19  
 21 2005). 21  
 23 2005). 23  
 25 2005). 25  
 27 2005). 27  
 29 2005). 29  
 31 2005). 31  
 33 2005). 33  
 35 2005). 35  
 37 2005). 37  
 39 2005). 39  
 41 2005). 41  
 43 2005). 43  
 45 2005). 45  
 47 2005). 47

7 Note that with natural satiety, ACh increases, 7  
 9 but DA is “normal” or elevated. However, during 9  
 11 drug withdrawal DA and ACh often respond in 11  
 13 opposite directions, with a decrease in DA and 13  
 15 increase in ACh. It was hypothesized that this im- 15  
 17 balance probably gives rise to an aversive state. 17  
 19 17  
 21 19  
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 47 45

## IV. Natural and artificial rewards: do they share common reward mechanisms and circuitry?

### IV.A. Sugar addiction

1 Natural reward mechanisms presumably were se- 1  
 3 lected in the wild to promote behaviors that are 3  
 5 necessary for the survival of the species. In normal 5  
 7 animals these reinforcers have opposing mecha- 7  
 9 nisms that inhibit the behavior once the specific 9  
 11 need has been satisfied. However, drugs of abuse 11  
 13 activate the reward circuits without necessarily ac- 13  
 15 tivating the inhibitory components of the regula- 15  
 17 tory system. A large body of evidence suggests that 17  
 19 natural reinforcers and drugs of abuse share com- 19  
 21 mon reward circuitry. For example, both food and 21  
 23 drug reinforcers increase extracellular DA in the 23  
 25 NAc (Di Chiara and Imperato, 1988; Hernandez 25  
 27 and Hoebel, 1988b; Radhakishun et al., 1988; 27  
 29 Pothos et al., 1991; Rada et al., 1991a; Salamone, 29  
 31 1994; Wise et al., 1995a, b; Tanda and Di Chiara, 31  
 33 1998; Cappendijk et al., 1999; Acquas et al., 2002). 33  
 35 These reinforcers seem to also share behavioral 35  
 37 responses. For instance, sweet taste or morphine 37  
 39 prolongs a meal. This can be blocked with nalox- 39  
 41 one, an opiate antagonist (Sclafani et al., 1982; 41  
 43 Nader et al., 1994; Gosnell et al., 1996). Con- 43  
 45 sumption of sugar can act as an analgesic by re- 45  
 47 leasing endogenous opioids (Kanarek et al., 1991). 47  
 Weight loss increases opiate-induced eating and 1  
 also drug self-administration (Hagan and Moss, 3  
 1991; Specker et al., 1994; Cabeza de Vaca and 5  
 Carr, 1998). 7

1 An animal model of binge eating has been developed by the Hoebel laboratory to systematically  
 3 study whether excessive sugar intake can elicit behavioral and neurochemical changes similar to  
 5 those of drugs of abuse (Avena et al., in press(a)). Several diagnostic criteria used to study drug  
 7 abuse reveal that binge eating of palatable foods, a major behavioral component of obesity, may have  
 9 some addictive-like properties. For example, rats maintained on a diet of intermittent access to a  
 11 sugar solution and chow gradually escalate their intake of sugar over the course of one month and  
 13 “binge” on the sugar when it becomes available each day (Colantuoni et al., 2001). These animals  
 15 also have increased D1 and mu-opioid receptor binding, and D3 receptor mRNA in the NAc  
 17 (Bassareo and Di Chiara, 1997; Colantuoni et al., 2001; Spangler et al., 2004). Drugs of abuse are  
 19 known to repeatedly increase DA release in the NAc without habituation of the response as seen  
 21 with palatable food (Bassareo and Di Chiara, 1997). Sugar bingeing also repeatedly releases DA  
 23 in the NAc (Rada et al., 2005), similar to addictive drugs (Fig. 4). A similar result is obtained when  
 25 sugar-bingeing rats sham-feed during the binge, suggesting that the taste of sugar is sufficient to  
 27 release DA repeatedly in the NAc (Avena et al., in press(b)). Signs of withdrawal such as teeth chattering,  
 29 grooming, anxiety, depression, and distress

vocalization are found in sugar-bingeing rats. This is most noticeable when withdrawal is precipitated  
 by an opioid antagonist, suggesting that the endogenous opioid system is altered by the excessive  
 bingeing. During both naloxone precipitated and even during simple withdrawal of the sugar (spontaneous  
 withdrawal, unpublished observations) microdialysis reveals decreases in DA and increases ACh in the  
 NAc (Colantuoni et al., 2002), indicative of a drug-like withdrawal state (Pothos et al., 1991; Rada et al.,  
 1991, 1996). Craving-like behavior is seen in sugar-binge animals as they manifest increased intake  
 after 2 weeks of abstinence, known as a “deprivation effect” (Avena et al., 2005). After a month of  
 abstinence the animals respond more than before for cues previously associated with sugar (Grimm et al.,  
 2005).

Evidence of cross-sensitization between sugar bingeing and withdrawal-induced locomotion (Avena  
 and Hoebel, 2003), withdrawal-induced locomotion (Gosnell, 2005), or enhanced alcohol intake  
 (Avena et al., 2004) has been shown, suggesting that a common neural pathway, presumably DA,  
 mediates these behaviors. Thus, each of these similarities suggest that binge eating on sugar  
 results in a state qualitatively similar to drug abuse, and that this state persists and can foster  
 future intake of sugar or drugs of abuse and suggests that a natural reinforcer such as sugar can

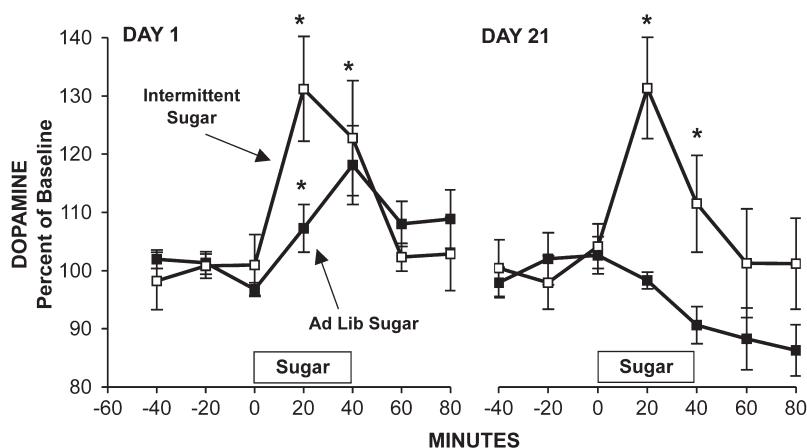


Fig. 4. Dopamine increases the first time rats have access to sugar; however, this response habituates and disappears in following trials if rats have *ad libitum* access to the sugar. In contrast, rats receiving intermittent sugar show the same DA response every time, similar to drugs of addiction.

change from a substance of use to a substance of abuse. All of these results point to a natural function for addiction that is usurped by the more powerful drugs of abuse.

## V. Conclusions

Brain microdialysis was invented by Ungerstedt (1984) and continues to be an immensely valuable technique for monitoring biochemicals and their metabolites in vivo. The measurement of release is one of the criteria for proving that a substance is a neurotransmitter. Moreover, measurement of release during behavior provides a critical piece of information in determining when and where the neurotransmitter-coded system is active. As faster time-sampling techniques are invented to match the rate of nerve impulse flow, microdialysis will continue to have a place for verifying measurement of the actual neurotransmitter in relation to behavior and underlying neural processes that depend on volume conduction through the extracellular and ventricular fluids of the brain.

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